

Biotransformation of Vermitaline by *Cunninghamella echinulata*

by Yi-Feng Lü^{a)}), Ke-Yu Chen^{a)}), Hui-Liang Li^{*a)}), Yue-Hu Pei^{b)}), Run-Hui Liu^{a)}),
Wei-Dong Zhang^{*a)c)})

^{a)} School of Pharmacy, Second Military Medical University, Shanghai 200433, P. R. China
(phone: +86-21-25070387; fax: +86-21-25070387; e-mail: faranli@hotmail.com)

^{b)} School of Traditional Chinese Materia Medica 49#, Department of Natural Medicinal Chemistry,
Shenyang Pharmaceutical University, Wenhua Road 103, Shenyang 110016, P. R. China

^{c)} School of Pharmacy, Shanghai Jiao Tong University, Shanghai 200240, P. R. China
(phone: +86-21-25070386; fax: +86-21-25070386; e-mail: wdzhangy@hotmail.com)

Biotransformation of vermitaline (**1**) by *Cunninghamella echinulata* (ATCC 30369) was carried out. Four biotransformation products were obtained and three of them were characterized as new compounds. On the basis of their NMR and mass-spectral data, their structures were characterized as 7 α -hydroxyrubijervine (**2**), 7 α -hydroxyrubijervine-7-O- β -D-galactofuranoside (**3**), 7 α -hydroxyvermitaline (**4**), and 7 α -hydroxyvermitaline-7-O- β -D-galactofuranoside (**5**).

Introduction. – Microbial transformation is an effective tool for the structural modification of bioactive natural compounds. Its application in asymmetric synthesis is increasing due to its versatility and ease [1]. A variety of transformations on natural products such as oxidation, reduction, hydrolysis, isomerization, epimerization, rearrangement, D-homoannulation, *Michael* addition, and reverse aldol reaction can be performed [2]. *Cunninghamella echinulata*, a filamentous fungus, is recognized for its potential for steroid hydroxylation and has been noted for its ability to mimic mammalian hepatic metabolism with other substrates [3][4].

Veratrum alkaloids are a group of potent hypotensive agents that act by reflex suppression of the cardiovascular system [5][6]. Vermitaline¹⁾ (**1**), a verazine type steroidal alkaloid, is one of the most extensively studied. The interest in *Veratrum* alkaloids has recently been renewed as they were patented several times during the past few years [7–9]. The potent regulatory effects of jervane *Veratrum* alkaloids on hedgehog signaling, modulation of cholesterol biosynthesis and transport, and control of cell proliferation during mandibular arch morphogenesis have been reported recently [10]. Biotransformation of *Veratrum* alkaloids, such as rubijervine, jervine and veratramine by various microorganisms has been reported previously [11–13].

In continuation of our studies on the phytochemistry of *Veratrum* alkaloids [14][15], we now report the characterization of four metabolites of vermitaline (**1**) in cell suspension of *C. echinulata* (ATCC 30369). The metabolites, which were more polar than **1**, were identified as 7 α -hydroxyrubijervine (**2**)¹⁾, 7 α -hydroxyrubijervine-7-O- β -D-galactoside (**3**)¹⁾, 7 α -hydroxyvermitaline (**4**)¹⁾ and 7 α -hydroxyvermitaline-7-O- β -D-galactoside (**5**)¹⁾ by spectroscopic methods.

¹⁾ For systematic names, see *Exper. Part*.

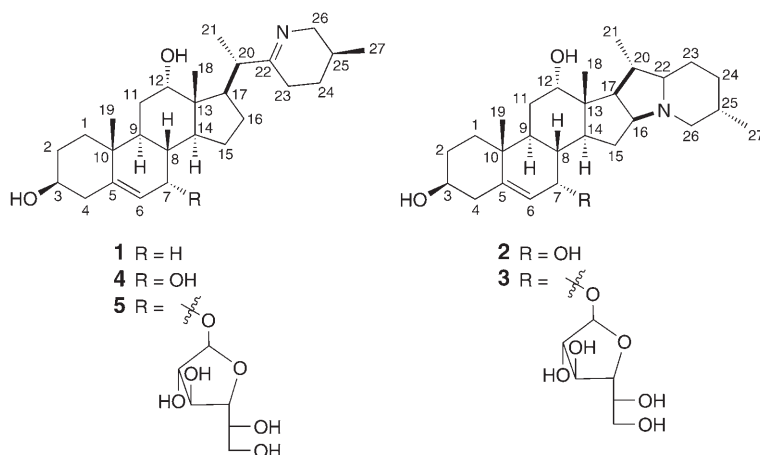


Figure. Structures of Compounds 1–5

Results and Discussion. – Compound **2** was obtained as a white amorphous powder, showing a positive reaction with *Dragendorff's* reagent. The HR-ESI-MS (positive mode) gave the molecular formula $C_{27}H_{43}NO_3$ from the $[M + H]^+$ signal at m/z 430.3314. The ESI-MS of **2** showed a significant fragment ion peak at m/z 412 ($[M + H - H_2O]^+$). The IR spectrum exhibited absorptions for OH groups (3370 and 1070 cm^{-1}). The ^{13}C -NMR spectrum (Table 1) of **2** showed signals due to four Me, eight CH_2 , and twelve CH groups, and three quaternary C-atoms, of which included one $\text{C}=\text{C}$ bond ($\delta(\text{C})$ 141.9 C(5), 122.3 C(6)) and three oxygenated sp^3 -C-atoms. The ^1H -NMR spectrum (Table 2) indicated H-atom resonances for four Me groups at δ 0.69 (s, Me(18)), 0.90 (d, $J = 6.0$, Me(21)), 0.96 (s, Me(19)), 0.97 (d, $J = 6.6$, Me(27)), and the ^{13}C -NMR exhibited the signals of three C-atoms attached to an N-atom at $\delta(\text{C})$ 73.7 (C(16)), 75.8 (C(22)), and 61.6 (C(26)), which are characteristic for a solanidine type steroidal alkaloid. Considering the steroidal alkaloids previously reported from the genus *Veratrum*, **2** was determined to be the known compound 7 α -hydroxyrubijervine¹) [16].

Compound **3** was obtained as a white amorphous powder, showing a positive reaction with *Dragendorff's* reagent. The HR-ESI-MS (positive mode) gave the molecular formula $C_{33}H_{53}NO_8$ from the $[M + H]^+$ signal at m/z 592.3842. The ESI-MS of **3** gave a significant fragment ion peak at m/z 430 ($[M + H - 162]^+$). The IR spectrum exhibited absorptions for OH groups (3361 and 1073 cm^{-1}). The ^{13}C -NMR data of **3** were similar to those of 7 α -hydroxyrubijervine (**2**), except for the presence of six signals at $\delta(\text{C})$ 60.8, 68.7, 75.8, 77.0, 80.3 and 101.9. From the above analysis, and considering the elimination of the 162 Da neutral fragment in the ESI-MS, we concluded that there is a β -galactose moiety (β -configured) in **3** by comparison of chemical shifts and coupling constants of the sugar moiety with those of galactofuranose [17], but the absolute configuration of the sugar units have not been proved. Therefore, **3** was determined as an analog of 7 α -hydroxyrubijervine (**2**). The galactofuranose moiety must be attached to C(7) due to the HMBC correlations

Table 1. ^{13}C -NMR Spectral Data of **1**–**5**. At 125 MHz in MeOH; δ in ppm.

	1	2	3	4	5
CH ₂ (1)	38.2	38.1	38.1	38.2	38.4
CH ₂ (2)	32.6	32.5	32.5	32.6	32.4
H–C(3)	71.9	72.1	70.2	71.9	72.1
CH ₂ (4)	43.0	42.8	42.3	43.0	43.1
C(5)	142.1	141.9	143.2	142.1	143.2
H–C(6)	122.4	122.3	124.5	122.4	122.7
CH ₂ (7) or H–C(7)	30.2	69.9	73.8	70.1	73.9
H–C(8)	37.0	36.8	38.2	37.0	39.4
H–C(9)	47.6	47.9	47.7	47.6	47.9
C(10)	34.8	34.9	34.8	34.8	35.0
CH ₂ (11)	33.3	32.9	31.6	33.3	32.9
H–C(12)	72.5	72.4	72.4	72.5	74.3
C(13)	48.3	48.5	48.3	48.3	48.6
H–C(14)	48.2	48.1	48.2	48.2	48.4
CH ₂ (15)	24.3	24.1	24.0	24.3	23.9
CH ₂ (16) or H–C(16)	27.4	73.7	72.8	27.5	27.3
H–C(17)	47.3	47.9	47.8	47.3	47.3
Me(18)	13.1	12.5	13.0	13.1	12.7
Me(19)	19.6	19.6	19.7	19.6	19.8
H–C(20)	47.8	47.8	47.6	47.8	47.9
Me(21)	17.6	17.6	17.6	17.6	17.6
C(22) or H–C(22)	179.1	75.8	75.9	179.1	179.1
CH ₂ (23)	28.0	28.2	28.4	28.0	28.1
CH ₂ (24)	28.2	28.5	28.5	28.2	28.2
H–C(25)	27.3	27.3	27.5	27.3	27.1
CH ₂ (26)	56.7	61.6	61.8	56.7	56.8
Me(27)	19.5	19.9	20.1	19.5	19.6
H–C(1')			101.9		102.0
H–C(2')			77.0		77.1
H–C(3')			75.8		76.0
H–C(4')			80.3		81.9
H–C(5')			68.7		69.9
CH ₂ (6')			60.8		61.9

between H–C(1') (5.09, *d*, $J = 7.0$) and C(7) ($\delta(\text{C})$ 73.8). Therefore, the structure of **3** was identified as 7 α -hydroxyrubijervine-7-O- β -D-galactofuranoside¹).

Compound **4** was obtained as a white amorphous powder, showing a positive reaction with *Dragendorff's* reagent. The HR-ESI-MS (positive mode) gave the molecular formula C₂₇H₄₄NO₃ from the $[M + \text{H}]^+$ signal at m/z 442.3304). The ESI-MS of **4** showed significant fragment ion peak at m/z 424 ($[M + \text{H} - \text{H}_2\text{O}]^+$). The IR spectrum exhibited absorptions for OH groups (3361 and 1068 cm⁻¹) and a C=C bond (1624 cm⁻¹). The ^{13}C -NMR (*Table 1*), ^1H -NMR (*Table 2*) and ESI-MS data suggested that **4** is a monohydroxylated vermitaline derivative [4]. Three OH groups were located at C(3) ($\delta(\text{C})$ 71.9), C(7) ($\delta(\text{C})$ 70.1), and C(12) ($\delta(\text{C})$ 72.5) respectively, due to the HMBC correlations between $\delta(\text{H})$ 3.20–3.24 (*m*, H–C(3)) and C(2) ($\delta(\text{C})$ 32.6), and C(4) ($\delta(\text{C})$ 43.0), $\delta(\text{H})$ 3.68 (*dd*, $J = 2.0, 2.0$, H–C(7)) and C(6) ($\delta(\text{C})$ 122.4), and C(8) ($\delta(\text{C})$ 37.0), $\delta(\text{H})$ 3.37 (*br. s*, H–C(12)) and C(11) ($\delta(\text{C})$ 33.3), and C(13) ($\delta(\text{C})$

Table 2. ¹H-NMR Spectral Data of **1–5**. At 500 MHz in MeOH; δ in ppm, *J* in Hz.

	1	2	3	4	5
CH ₂ (1)	1.53–1.55 (<i>m</i>), 1.48–1.52 (<i>m</i>)	1.54–1.57 (<i>m</i>), 1.48–1.53 (<i>m</i>)	1.53–1.56 (<i>m</i>), 1.47–1.52 (<i>m</i>)	1.53–1.56 (<i>m</i>), 1.49–1.52 (<i>m</i>)	1.53–1.57 (<i>m</i>), 1.48–1.59 (<i>m</i>)
CH ₂ (2)	1.60–1.64 (<i>m</i>), 1.30–1.34 (<i>m</i>)	1.60–1.63 (<i>m</i>), 1.30–1.35 (<i>m</i>)	1.59–1.64 (<i>m</i>), 1.32–1.36 (<i>m</i>)	1.62–1.64 (<i>m</i>), 1.31–1.35 (<i>m</i>)	1.58–1.62 (<i>m</i>), 1.30–1.34 (<i>m</i>)
H–C(3)	3.22–3.26 (<i>m</i>)	3.21–3.26 (<i>m</i>)	3.22–3.25 (<i>m</i>)	3.20–3.24 (<i>m</i>)	3.21–3.25 (<i>m</i>)
CH ₂ (4)	2.12–2.15 (<i>m</i>), 2.08–2.11 (<i>m</i>)	2.11–2.15 (<i>m</i>), 2.07–2.10 (<i>m</i>)	2.13–2.16 (<i>m</i>), 2.09–2.11 (<i>m</i>)	2.13–2.16 (<i>m</i>), 2.08–2.13 (<i>m</i>)	2.13–2.16 (<i>m</i>), 2.08–2.12 (<i>m</i>)
H–C(6)	5.35 (<i>d</i> , <i>J</i> =2.0)	5.35 (<i>d</i> , <i>J</i> =2.0)	5.32 (<i>d</i> , <i>J</i> =2.0)	5.32 (<i>d</i> , <i>J</i> =2.0)	5.32 (<i>d</i> , <i>J</i> =2.0)
CH ₂ (7) or H–C(7)	3.68 (<i>dd</i> , <i>J</i> =2.0, 2.0)	3.68 (<i>dd</i> , <i>J</i> =2.0, 2.0)	3.30 (<i>dd</i> , <i>J</i> =2.0, 2.0)	3.68 (<i>dd</i> , <i>J</i> =2.0, 2.0)	3.30 (<i>dd</i> , <i>J</i> =2.0, 2.0)
H–C(8)	1.43–1.47 (<i>m</i>)	1.43–1.47 (<i>m</i>)	1.75–1.79 (<i>m</i>)	1.45–1.49 (<i>m</i>)	1.65–1.69 (<i>m</i>)
H–C(9)	2.10–2.12 (<i>m</i>)	2.08–2.12 (<i>m</i>)	2.08–2.13 (<i>m</i>)	2.09–2.12 (<i>m</i>)	2.07–2.10 (<i>m</i>)
CH ₂ (11)	1.66–1.70 (<i>m</i>), 1.43–1.47 (<i>m</i>)	1.65–1.69 (<i>m</i>), 1.44–1.47 (<i>m</i>)	1.67–1.70 (<i>m</i>), 1.43–1.46 (<i>m</i>)	1.65–1.08 (<i>m</i>), 1.43–1.47 (<i>m</i>)	1.66–1.70 (<i>m</i>), 1.45–1.48 (<i>m</i>)
H–C(12)	3.50 (<i>br. s</i>)	3.50 (<i>br. s</i>)	3.37 (<i>br. s</i>)	3.37 (<i>br. s</i>)	3.37 (<i>br. s</i>)
H–C(14)	2.08–2.12 (<i>m</i>)	2.07–2.12 (<i>m</i>)	2.09–2.13 (<i>m</i>)	2.08–2.11 (<i>m</i>)	2.08–2.12 (<i>m</i>)
CH ₂ (15)	1.85–1.87 (<i>m</i>), 1.09–1.13 (<i>m</i>)	1.85–1.89 (<i>m</i>), 1.09–1.12 (<i>m</i>)	1.84–1.87 (<i>m</i>), 1.08–1.11 (<i>m</i>)	1.83–1.87 (<i>m</i>), 1.09–1.13 (<i>m</i>)	1.85–1.88 (<i>m</i>), 1.10–1.13 (<i>m</i>)
CH ₂ (16) or H–C(16)	1.33–1.36 (<i>m</i>), 1.58–1.61 (<i>m</i>)	2.21–2.24 (<i>m</i>)	2.21–2.24 (<i>m</i>)	1.34–1.36 (<i>m</i>), 1.58–1.62 (<i>m</i>)	1.33–1.37 (<i>m</i>), 1.57–1.61 (<i>m</i>)
H–C(17)	2.24 (<i>dd</i> , <i>J</i> =10.0, 10.0)	2.24 (<i>dd</i> , <i>J</i> =10.0, 10.0)	2.24 (<i>dd</i> , <i>J</i> =10.0, 10.0)	1.50–1.53 (<i>m</i>)	1.50–1.53 (<i>m</i>)
Me(18)	0.69 (<i>s</i>)	0.69 (<i>s</i>)	0.69 (<i>s</i>)	0.76 (<i>s</i>)	0.76 (<i>s</i>)
Me(19)	0.96 (<i>s</i>)	0.96 (<i>s</i>)	0.96 (<i>s</i>)	1.01 (<i>s</i>)	1.01 (<i>s</i>)
H–C(20)	1.63–1.67 (<i>m</i>)	1.63–1.66 (<i>m</i>)	1.64–1.67 (<i>m</i>)	1.88–1.91 (<i>m</i>)	1.88–1.92 (<i>m</i>)
Me(21)	0.90 (<i>d</i> , <i>J</i> =6.0)	0.90 (<i>d</i> , <i>J</i> =6.0)	0.90 (<i>d</i> , <i>J</i> =6.0)	1.05 (<i>d</i> , <i>J</i> =5.0)	1.05 (<i>d</i> , <i>J</i> =5.0)
H–C(22)	1.48–1.52 (<i>m</i>)	1.49–1.52 (<i>m</i>)	1.48–1.53 (<i>m</i>)	–	–
CH ₂ (23)	1.64–1.68 (<i>m</i>), 1.20–1.23 (<i>m</i>)	1.63–1.66 (<i>m</i>), 1.21–1.25 (<i>m</i>)	1.62–1.66 (<i>m</i>), 1.20–1.24 (<i>m</i>)	2.33–2.37 (<i>m</i>), 2.23–2.27 (<i>m</i>)	2.34–2.37 (<i>m</i>), 2.23–2.26 (<i>m</i>)
CH ₂ (24)	1.62–1.66 (<i>m</i>), 0.94–0.98 (<i>m</i>)	1.63–1.66 (<i>m</i>), 0.93–0.97 (<i>m</i>)	1.64–1.67 (<i>m</i>), 0.95–0.98 (<i>m</i>)	1.63–1.68 (<i>m</i>), 0.97–1.00 (<i>m</i>)	1.62–1.65 (<i>m</i>), 0.94–0.99 (<i>m</i>)
H–C(25)	1.47–1.51 (<i>m</i>)	1.49–1.52 (<i>m</i>)	1.47–1.50 (<i>m</i>)	1.48–1.52 (<i>m</i>)	1.47–1.52 (<i>m</i>)
CH ₂ (26)	2.84 (<i>dd</i> , <i>J</i> =11.0, 3.5), 1.34 (<i>dd</i> , <i>J</i> =11.0, 3.5)	2.84 (<i>dd</i> , <i>J</i> =11.0, 3.5), 1.34 (<i>dd</i> , <i>J</i> =11.0, 3.5)	1.45 (<i>dd</i> , <i>J</i> =12.0, 6.0), 1.19 (<i>dd</i> , <i>J</i> =12.0, 5.8)	1.45 (<i>dd</i> , <i>J</i> =12.0, 6.0), 1.19 (<i>dd</i> , <i>J</i> =12.0, 5.8)	1.45 (<i>dd</i> , <i>J</i> =12.0, 6.0), 1.19 (<i>dd</i> , <i>J</i> =12.0, 5.8)
Me(27)	0.95 (<i>d</i> , <i>J</i> =6.6)	0.97 (<i>d</i> , <i>J</i> =6.6)	0.95 (<i>d</i> , <i>J</i> =6.0)	0.94 (<i>d</i> , <i>J</i> =6.0)	0.97 (<i>d</i> , <i>J</i> =6.0)
H–C(1')			5.09 (<i>d</i> , <i>J</i> =7.0)		5.09 (<i>d</i> , <i>J</i> =7.0)
H–C(2')			3.96–3.98 (<i>m</i>)		3.96–3.98 (<i>m</i>)
H–C(3')			4.51–4.53 (<i>m</i>)		4.51–5.54 (<i>m</i>)
H–C(4')			3.91–3.93 (<i>m</i>)		3.90–3.93 (<i>m</i>)
H–C(5')			3.62–3.65 (<i>m</i>)		3.62–3.65 (<i>m</i>)
CH ₂ (6')			3.81 (<i>dd</i> , <i>J</i> =2.0, 3.0), 3.79 (<i>dd</i> , <i>J</i> =2.0, 3.0)		3.81 (<i>dd</i> , <i>J</i> =2.0, 3.0), 3.79 (<i>dd</i> , <i>J</i> =2.0, 3.0)

48.3). The configuration of the positions C(3), C(12), C(13), C(10), C(20), and C(25), respectively, was established to be analogous to **1** by analysis of NOESY spectra, and also by comparison of chemical shifts and coupling constants of **4** with those of **1**. The relative configuration of C(7) was deduced to be α , by the observation of the NOESY correlation between H–C(7) with $\delta(\text{H})$ 1.45–1.49 (m , H_β –C(8)). Therefore, the structure of **4** was identified as 7 α -hydroxyvermitaline¹).

Compound **5** was obtained as a white amorphous powder, showing a positive reaction with *Dragendorff's* reagent. The HR-ESI-MS (positive mode) of **5** displayed a peak at m/z 592.3853 ($[M+H]^+$), suggesting the molecular formula $\text{C}_{33}\text{H}_{53}\text{NO}_8$. The IR spectrum exhibited absorptions for OH groups (3352 and 1068 cm^{-1}) and C=C bonds (1618 cm^{-1}). The ^{13}C -NMR (Table 1), and ^1H -NMR (Table 2) spectra of **5** suggested the structure of a vermitaline derivative. Similar to compound **3**, the ^{13}C -NMR spectrum of **5** showed six signals in the region at $\delta(\text{C})$ 61.9, 69.9, 76.0, 77.1, 81.9, and 102.0. These data indicated the presence of a β -D-galactose moiety in **5**. H–C(7) ($\delta(\text{H})$ 3.30) also showed HMBC coupling with C(1') ($\delta(\text{C})$ 102.0). Metabolite **5** was thus confirmed to be 7 α -hydroxyvermitaline-7-O- β -D-galactofuranoside¹), a new metabolic product reported for the first time in fermentation studies of steroidal alkaloids.

Experimental Part

General. Vermitaline (= (3 β ,12 α ,17 β)-17-[1-[(5S)-3,4,5,6-tetrahydro-5-methylpyridin-2-yl]ethyl]androst-5-ene-3,12-diol; **1**) was isolated from the roots of *Veratrum dahuricum* with a purity of 99.1% by HPLC analysis. The compound was authenticated by detailed NMR analysis and comparing with data from the literature [18]. Column chromatography (CC): MCI gel CHP20P (high-porous polymer 75–150 μ ; Mitsubishi Chemicals, Japan), reversed-phase silica gel RP-18 (40–63 μm , Merck, Germany), silica gel (SiO_2) (200–300 mesh, Yantai, P. R. China) and Sephadex LH-20 (Pharmacia Co. Ltd.). TLC: SiO_2 plates; visualization by spraying with 10% H_2SO_4 in EtOH, followed by heating. Optical rotations: Perkin-Elmer-341 polarimeter. IR Spectra: Nicolet-Compact-40-Bruker-Vector-22 spectrophotometer; ν_{max} in cm^{-1} . NMR Spectra: Bruker AVANCE^{II} 500 NMR, for ^1H -NMR at 500 MHz, and ^{13}C -NMR at 125 MHz; δ in ppm with Me_4Si as an internal standard, J in Hz. MS: Varian MAT-212 mass spectrometer (for ESI) and Q-Tof micro mass spectrometer (for HR-ESI); in m/z .

Plant Material. The plants were collected in Yanbian, Jilin Province, P. R. China, in September 2005, and authenticated as *V. dahuricum* by Prof. Yong-Zhen Liu, School of Pharmacy, Yanbian University, China. A voucher specimen (No. 211) was deposited at the Herbarium of the School of Pharmacy, Shanghai Jiao Tong University, Shanghai, China.

Organisms. The stock culture of *C. echinulata* (ACCC 30369) was maintained on a potato dextrose agar slant. Four Erlenmeyer flasks (1000 ml), each containing 300 ml of liquid medium consisting of 0.1% peptone, 0.1% yeast extract, 0.1% beef extract, and 0.5% glucose were inoculated with freshly obtained *C. echinulata* cultured from the agar slant on a rotary shaker at 180 rpm. After cultivation at 28° for 72 h, the vermitaline (**1**) soln. (50 mg of vermitaline dissolved in 250 μl EtOH) was added to each flask, and the incubation continued for 12 d.

Microbial Metabolism of Vermitaline. After 12 d of incubation, the incubation mixtures were pooled and filtered. The filtrate was applied to a MCI-gel CHP20P column, and washed with H_2O to remove sugars, then it was eluted with 20% aq. MeOH, 60% aq. MeOH, and MeOH to give the fractions A, B, and C resp. Each fraction was evaporated to dryness *in vacuo* and analyzed by TLC, which was developed by $\text{CHCl}_3/\text{MeOH}$ (10:1) and visualized by spraying with 10% H_2SO_4 soln. Frs. B and C were combined and chromatographed repeatedly through a SiO_2 column (CHCl_3) and Sephadex LH-20 (70% MeOH) to furnish **2** (4.9 mg) and **4** (5.6 mg). Fr. A was dissolved in 10% aq. MeOH and subjected to RP SiO_2 (ODS) CC, eluting with the gradient $\text{H}_2\text{O}/\text{MeOH}$ (100:10–0:100) to give **3** (5.9 mg), and **5** (3.8 mg).

7 α -Hydroxyrubijervine (= (3 β ,7 α ,12 α ,22 ξ)-Solaniid-5-ene-3,7,12-triol; **2**). White amorphous powder. IR (KBr): 3370, 2951, 2177, 1623, 1070. ¹H- and ¹³C-NMR: *Tables 1* and 2. 2D-NMR (HMBC; 500 MHz, CDCl₃): H–C(3)/C(4), H–C(7)/C(5), H–C(7)/C(8), H–C(15)/C(14). ESI-MS: 430 ([M + H]⁺), 412 ([M + H – H₂O]⁺). HR-ESI-MS: 430.3314 ([M + H]⁺, C₂₇H₄₄NO₃⁺; calc. 430.3321).

7 α -Hydroxyrubijervine-7-O- β -D-galactofuranoside (= (3 β ,7 α ,12 α ,22 ξ)-3,12-Dihydroxysolaniid-5-en-7-yl Hexofuranoside; **3**). White amorphous powder. IR (KBr): 3361, 2947, 2170, 1617, 1073. ¹H- and ¹³C-NMR: *Tables 1* and 2. 2D-NMR (HMBC; 500 MHz, CDCl₃): H–C(3)/C(4), H–C(7)/C(5), H–C(7)/C(8), H–C(15)/C(14), H–C(1')/C(7). ESI-MS: 592 ([M + H]⁺), 430 ([M + H – 162]⁺). HR-ESI-MS: 592.3842 ([M + H]⁺, C₃₃H₅₄NO₈⁺; calc. 592.3849).

7 α -Hydroxyvermitaline (= (3 β ,7 α ,12 α ,17 β)-17-[1-[(5S)-3,4,5,6-Tetrahydro-5-methyl-2-pyridinyl]ethyl]andro-5-ene-3,7,12-triol; **4**). White amorphous powder. IR(KBr): 3361, 2937, 2155, 1624, 1068. ¹H- and ¹³C-NMR: *Tables 1* and 2. 2D-NMR (HMBC; 500 MHz, CDCl₃): H–C(3)/C(4), H–C(7)/C(5), H–C(7)/C(8), H–C(15)/C(14). ESI-MS: 430 ([M + H]⁺), 658 ([M + H – H₂O]⁺). HR-ESI-TOF-MS: 430.3304 ([M + H]⁺, C₂₇H₄₄NO₃⁺; calc. 430.3321).

7 α -Hydroxyvermitaline-7-O- β -D-galactofuranoside (= (3 β ,7 α ,12 α ,17 β)-3,12-Dihydroxy-17-[1-[(5S)-3,4,5,6-tetrahydro-5-methyl-2-pyridinyl]ethyl]andro-5-en-7-yl Hexofuranoside; **5**). White amorphous powder. IR(KBr): 3352, 2944, 2162, 1618, 1068. ¹H- and ¹³C-NMR *Tables 1* and 2. 2D-NMR (HMBC; 500 MHz, CDCl₃): H–C(3)/C(4), H–C(7)/C(5), H–C(7)/C(8), H–C(15)/C(14), H–C(1')/C(7). ESI-MS: 592 ([M + H]⁺), 430 [M + H – 162]⁺. HR-ESI-MS: 592.3853 ([M + H]⁺, C₃₃H₅₄NO₈⁺; calc. 592.3849).

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